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# DIFFERENTIAL RESPONSE OF CITRUS GERMPLASM FOR VARYING CONCENTRATION OF ALUMINIUM

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### ABSTRACT

Acidic soils with high amount of aluminium are the major problems of hilly regions of India. A study was conducted to evaluate the effect of aluminium concentrations on the growth parameters viz. root growth, root volume, shoot growth, dry weight of citrus germplasm. Fourteen germplasm of citrus were grown in hydroponics in Hoagland's solution having different concentrations of aluminium with 0, 50, 100, 200 and 400  $\mu$ molL<sup>-1</sup> of Al<sub>2</sub> (SO<sub>4</sub>)<sub>3</sub>.18H<sub>2</sub>O. The results revealed that there was general reduction in all the growth parameters observed. In general, there was a difference of response in all the germplasm tested. Among the various growth parameters, the root length and root volume were reduced markedly by the increase in concentration of aluminium. Pani Jyamir (G<sub>10</sub>) and Citron (G<sub>14</sub>) can tolerate higher amount of Al toxicity which can be use as root stock or can be cultivated under Al toxic conditions and that of Nagpur mandarin (G<sub>5</sub>) was with highest reduction. Further Principal component analysis and cluster analysis were also used for determining the susceptible and tolerant germplasm for aluminium toxicity and also to group germplasm based on their tolerance on various Al concentrations.

*Keywords*: Acidic soil, aluminium toxicity, growth parameters, citrus germplasm, Hydroponics, multivariate analysis, screening, acidic soil.

#### Introduction

Acidic soils with high amount of aluminium are the major problems of hilly regions of India. Plants which are grown in acid soils, due to aluminium solubility at low pH have under developed root system and exhibit a variety of nutrient deficiency symptoms consequently decrease in yield. Al interferes with nutrients uptake, transport and utilization of essential nutrients including Ca, Mg, K, P, Cu, Fe, Mn and Zn (Foy, 1984; Guo et al., 2003). Making of successful citrus plantation in acidic soil will be mainly depend upon identifying the germplasms which are tolerant for aluminium toxicity either to be used as cultivar or to be used as rootstocks. Since India has large number of variation in citrus germplasm screening them for aluminium toxicity will be a better option. Hence, a study was conducted to test the effect of varying concentrations of aluminium on the growth parameters of different citrus germplasm.

#### **Materials and Methods**

The research was carried out at green house of Department of Horticulture, Sikkim University, Gangtok, during the period of five months i.e., March-December 2014 at an elevation of 1230 m above M.S.L with latitude and longitude of N 27'18.495' and E 88' 35. 307' respectively. The fruits were obtained from Sikkim i.e., from Majhitar Horticulture Farm, KVK East, Martam Nazitam and outside Sikkim i.e., from Nagpur, Maharastra and Pasighat, Arunachal Pradesh which were tagged separately and were washed, demucilaged and then seeds were extracted using running water. The seeds were dried separately in room temperature for a day and were kept in a separate folded paper bag.

Seeds were sown in germination sand within a week of extraction in a utility tray of size 36.83cm X 25.4cm with 14 varieties @ 10 seeds per variety in each tray. They were sprayed with Hoagland's solution

and distilled water till the germination and transplantation. After germination the germplasms were transplanted to hydroponic system in the trays containing 4 litre of double distilled water and one litre of Hoagland solution which was modified according to Simon *et al.* (1994). The seedlings were transplanted on 56<sup>th</sup> day from sowing. All the germplasms were transplanted to five different trays along with their replications.

Each tray containing Hoagland's solution and aluminium was aerated by means of air compressor. Macronutrients were added as  $PO4^{2-}$  (KH<sub>2</sub>PO<sub>4</sub>), 3.38 cmol<sub>c</sub> kg<sup>-1</sup>., K<sup>+</sup> (KNO<sub>3</sub>), 9,00 cmol<sub>c</sub> kg<sup>-1</sup>., Ca<sup>2+</sup> [Ca(NO<sub>3</sub>)<sub>2</sub>.H<sub>2</sub>O],10.13 cmol<sub>c</sub> kg<sup>-1</sup>., Mg<sup>2+</sup> (MgSO<sub>4</sub>. 7H<sub>2</sub>O), 3.38 cmol<sub>c</sub> kg<sup>-1</sup> while the micronutrients were added as Fe<sup>2+</sup> (FeCl<sub>3</sub> 6H<sub>2</sub>O), 1.6 mgL<sup>-1</sup>., Na<sup>+</sup> (Na<sub>2</sub> EDTA.H<sub>2</sub>O)., 1.43 mgL<sup>-1</sup>., Mn<sup>3+</sup> (MnSO<sub>4</sub>. 4H<sub>2</sub>O)., 0.25 mg L<sup>-1</sup>., MoO<sub>4</sub> [(NH<sub>4</sub>)6Mo<sub>7</sub>O<sub>24</sub>. 4H<sub>2</sub>O],0.006 mgL<sup>-1</sup>., B (H<sub>3</sub>BO<sub>3</sub>),0.37 mgL<sup>-1</sup>., Zn<sup>2+</sup> (ZnSO<sub>4</sub>. 7H<sub>2</sub>O) ,0.12mgL<sup>-1</sup> and Cu<sup>2+</sup> (CuSO<sub>4</sub>.5H<sub>2</sub>O), 0.03 mgL<sup>-1</sup>.

After seven days of seedling establishment in the trays, each tray was added five different concentrations of aluminium as 0, 50, 100, 200 and 400  $\mu$ mol L<sup>-1</sup>. The pH of the solution was monitored daily and maintained at 4.0 by adding 0.1 mol L<sup>-1</sup> HCl or NaOH solutions, as necessary. The solution was changed in every two weeks.

After 60 days of growth the germplasms were removed from hydroponics system and subjected to the growth parameters analysis. Root and shoot measurement as described by Lin and Myhre, 1991, entire plant dry weight as per Pereira *et al.* (2003) and root volume by water displacement method was carried out. The root and shoot length were measured in centimetre scale. The experimental design was carried with Factorial Completely Randomized Design (CRD).

Each concentration of aluminium was treated as one treatment (Factor-1). Under each treatment fourteen germplasms (Factor-2) were screened and for each germplasm three replications were used. Statistical analysis was carried out by one-way ANOVA (analysis of variance) to compare the significance of difference between the treatments and germplasms.

#### **Results and Discussion**

#### Morphological characters

#### Plant height (cm)

Plant height of *Citrus sps.* was measured in five weeks intervals. It was found to be significantly different for all the germplasms, aluminium concentration in different levels and their interaction

effect.  $G_1$  was found to be the tallest among all the germplasm under various aluminium concentrations in fifth, fourth, third, second and first week of growth (table). In addition, fifth, fourth, second and first week of planting the growth declined at  $A_{50}$ . Whereas, fifth, fourth, second and  $1^{st}$  week of planting at  $A_{100}$  the height increased slowly.

In this study it was analysed that at  $A_{400}$  (highest aluminium concentration), the height of plant in  $1^{st}$ , third, fourth and fifth week of planting except at  $2^{nd}$  week where there was slight decline (table 3). The interaction effect between genotype and aluminium level showed highest reduction plant height at fifth, fourth and second week of planting.

#### Root length (cm)

Among all the fourteen-germplasm tested, Nagpur mandarin showed highest reduction of root growth. On the other hand, increase of root growth was observed in three different germplasms namely Rangpur lime  $(G_9)$ , Citron  $(G_{14})$  and Pani Jyamir  $(G_{10})$ . Infact Rangpur lime  $(G_9)$  showed significant increase in root length.

From the root length analysis, it was found that Rangpur lime  $(G_9)$  was highly tolerant to Al toxicity followed by citron  $(G_{14})$ , Pani Jyamir  $(G_{10})$ , Sour orange  $(G_6)$ , Mosambi  $(G_7)$ , Pomelo  $(G_{12})$  and Sikkim mandarin  $(G_2)$ . The highly susceptible species were Nagpur mandarin  $(G_5)$  followed by Indian mandarin  $(G_1)$ , Grape fruit  $(G_{11})$ , Cleopatra mandarin  $(G_8)$ , Rough lemon  $(G_3)$  and Acid lime  $(G_4)$  as illustrated in Table 3 and Fig. 1a, 1b and 1c

This trend was similar to the earlier report by Yokomizo and Ishihara (1973) in Natsudaidai seedlings; Lin and Myhre (1991) in Carrizo citrange, Cleopatra mandarin, rough lemon, Sour orange and swingle citrumelo; Santos *et al.* (1999) in Rangpur lime and swingle citrumelo; Pereira *et al.* (2003) in rough lemon and Cleopatra mandarin and Batista *et al.* (2013) in corn plants who mentioned significant reduction in root length with increasing Al concentrations.

#### Volume of root (cc)

The result presented on aspect of volume of root showed decreasing trend with increase in concentration of aluminium in several germplasms (Table 4 and Fig. 2.) Highest reduction was in Nagpur mandarin  $(G_5)$  whereas increase in volume of root compared to control was found in Sikkim mandarin  $(G_2)$ , Rough lemon  $(G_3)$ , Acid lime  $(G_4)$ , Sour orange  $(G_6)$ , Mosambi  $(G_7)$  and Citron  $(G_{14})$ . However, among the germplasm, Mosambi  $(G_7)$  showed highest significant

increase in root volume compared to control, which is reported for the first time in present study.

From the root volume analysis, it was evident that *Citrus sinensis* ( $G_7$ ) was found most tolerant to root volume followed by Sour orange ( $G_6$ ) and Acid lime ( $G_4$ ). On the other hand, highly, susceptible species were: Nagpur mandarin ( $G_5$ ), Cleopatra mandarin ( $G_8$ ), Pomelo ( $G_{12}$ ) and Kala Jyamir ( $G_{12}$ ).

The trend was in corroboration with the findings of Santos *et al.* (1999a) who mentioned that increase in aluminium concentration reduces the root volume showing inverse relationship of root volume and aluminium concentration. Their study revealed Swingle citrumelo and Rangpur lime with reduction in root volume at 7.5 mgL<sup>-1</sup>. Whereas in Cravo lemon there was no significant influence in the root volume at this concentration. However, at 30 mgL<sup>-1</sup> the decrease was observed.

#### Shoot length (cm)

In shoot length as depicted in Table 5 and Fig. 3, it was observed that several germplasms had shown reduction compared to control. Decrease in shoot length was recorded in eleven germplasm and that of increase in three germplasms which were significantly higher than control. Nagpur mandarin  $(G_5)$  showed highest reduction. On the other hand, increase in shoot height was found in three different germplasms *viz:* Pani Jyamir  $(G_{10})$ , Pomelo  $(G_{12})$  and Bimira  $(G_{14})$ . In addition, Pani Jyamir  $(G_{10})$  was among the fourteen germplasm which showed the best result in terms of tolerance to Al stress level and that of Nagpur mandarin  $(G_5)$  was observed with susceptibility.

Based on the shoot length analysis on Al stress condition, The sequential order of germplasm for Al tolerance can be Pani Jyamir  $(G_{10})$ >Pomelo  $(G_{12})$ > Citron  $(G_{14})$ >Rangpur lime  $(G_9)$ >Kala Jyamir  $(G_{13})$ >Sour orange  $(G_6)$ >Sikkim mandarin $(G_2)$ > Cleopatra mandarin  $(G_8)$ , Mosambi $(G_7)$ , Indian mandarin  $(G_1)$ > Grape fruit  $(G_{11})$ > Rough lemon  $(G_3)$ > Acid lime  $(G_4)$ > Nagpur mandarin  $(G_5)$ . The report is presented first time in this study.

#### Dry weight of plant (g)

Dry weight of plant (DWP) invariably reduced with respect to increase in aluminium stress levels. Out of the fourteen-germplasm tested reduction was found in twelve germplasm and that of increase in two germplasms compared to control. Among all the germplasm, Nagpur mandarin  $(G_5)$  showed highest reduction of dry weight of plant on the other hand increase of dry weight was in two germplasms Sikkim mandarin  $(G_2)$  and Pani Jyamir  $(G_{10})$ . Infact Sikkim mandarin  $(G_2)$  had shown significant increase in dry

weight of plant which is reported for the first time in this study.

From the dry weight of plant analysis, it was found that Sikkim mandarin  $(G_2)$  was most tolerant to aluminium toxicity followed by in most to least tolerance as Pani Jyamir  $(G_{10})$ , Acid lime  $(G_4)$ , Kala Jyamir  $(G_{13})$ , Rangpur lime  $(G_9)$ , Mosambi  $(G_7)$ , Sour orange  $(G_6)$ , Pomelo  $(G_{12})$ , Rough lemon  $(G_3)$ , Cleopatra mandarin  $(G_8)$ , Grape fruit  $(G_{11})$  and Indian mandarin  $(G_1)$  as mentioned in Table 6 and Fig 4.

The result obtained in the research is in consonance with the findings of Pereira *et al.* (2003). They had mentioned that increase in Al will reduce the dry matter of leaves. The similar kind of report was presented by Santos *et al.*(1999b) who mentioned reduction in dry weight of leaves, root, stem in Swingle citrumelo with increase in Al levels and that of dry weight of lemon carnation was not significantly influenced by different levels of Al. Further, Nogueira *et al.* (1989) reported in Rangpur lime, Sunki, Tangerine, tangelo Orlando, Volkamer lemon roots and leaves had shown decrease in their dry weight with increase in aluminium levels.

## Principal component analysis (PCA) and Cluster analysis

Based on the PCA, at control PC1 and PC2 contribute 82.17 % of the total variance within the dataset analysed. Root length (Rl), root volume (Rv) and shoot length (Sl) showed positive value for both PC1 and PC2. Whereas, PH1, PH2, PH3 and DWP were found with positive value for PC1 and PC2 with negative value (fig. 5). The PC analysis and cluster analysis resulted in four group of Citrus germplasm based on the resistance for the various levels of aluminium.

Group first had G1 and G11 with higher values for PH1, PH2, PH3 and DWP and moderate value of Rl, Rv and Sl. Germplasm G1, G3, G4 could be grouped in another group based on the high value of Rl, Rv and Sl and showed moderate values for remaining growth response under study. Third group comprised of G2, G8, G10 and G13 with moderate values for all the growth parameters under study. The fourth group consist of G14, G6, G9 and G7. Amongst all it had lower values for all the character under study.

At,  $A_{50} \,\mu \text{MolL}^{-1}$  grouped them into four groups by PC analysis and cluster analysis of the germplasm under study (Fig. 6). In this PC1 and PC2 contributed 87% of the total variance present in the dataset. At 50  $\mu \text{MolL}^{-1} \,\mu \text{M}$ , Rl, Rv, Sl and PH1 had positive value for PC1 and negative value for PC2 (fig.). The first group includes of G3, G10 and G4 which was found with high value for the parameters *viz:* Rl, Rv, Sl and PH1 and moderate value for characters *viz:* PH2, PH3, PH4

and DWP. Further, G1 and G5 could be considered as the second group had higher values characters for PH2, PH3, PH4 and DWP and moderate value for Rl, Rv, Sl and PH1. Likewise, germplasm, G2 grouped as the third group had moderate values for the characters under study.

At  $100~\mu MolL^{-1}$  of aluminium applied, PC1 and PC2 both contributed 83.4% of the total variance in the data set. Based on PC and cluster analysis, all the germplasm were grouped in four clusters (fig. 7). From the fig 7 it was revealed that Rl, Rv, DWP, PH1 showed positive values for both PC1 and PC2, while PH1, PH2 and Sl had positive values at PC1 and negative value at PC2. In the first group, G11, G4 and G7 had shown moderate values for all the parameters under the research.

Further germplasm, G3, G5, G9 and G13 could be placed in second group which had shown higher value for parameters like Rl, Rv, DWP and PH5 but was moderate level in PH1, PH2 and Sl. Likewise, G2, G4 and G7 was found in another group and the last group comprises of G5, G11, G12 and G6 was found with lower values for all the parameters under the research.

Moreover at 200 µMolL<sup>-1</sup> µM of aluminium level, four groups were categorised where PC1 and PC2 showed 88.9 % of the total variance of dataset. In both PC1 and PC2, parameters like Rv, DWP and PH2 showed positive response. But Rl and Sl had positive

value for PC1 and negative value for PC2 accordingly (fig 8). Germplasm G3, G10 and G12 were found in one group. Rv, DWP and Ph2 had shown higher response for this group of germplasm while R1 and S1 were moderate for this group. The second group was formed from G2, G4, G7 and G13 was found with moderate values for Rv, PH2 and DWP and lower values for Rl and S1 which can be grouped in another group accordingly. Finally, the fourth group consist of G9, G8 which have lesser value for all the parameters and can be grouped into one group.

Last group (400 µMolL<sup>-1</sup> of aluminium), which was the highest dose of aluminium applied was found with 77.4% of the total variance in the dataset. RI, SI, DWP and PHP were the parameters which is found in G8, G9, G10, G11 and G12. In addition, PH1, PH2 and Rv were higher in G1. G2, G4 and G14 were on the third group and lastly G3, G5, G6, G7 and G13 were i the last group which had moderate of all the parameters.

#### Conclusion

Among the germplasm used in the experiment, Pani Jyamir and Citron were found to be tolerant and Nagpur Mandarin was found to be highly susceptible for aluminium toxicity. Pani Jyamir, a strain of *Citrus jambhiri* and Citron (*Citrus medica*) can be used a root stock for tolerance to aluminium toxicity.

**Table 1:** Germplasm used for Experiment

Sl. No.	Common name	Scientific Name
$G_1$	Indian Mandarin	Citrus indica
$G_2$	Sikkim Mandarin	Citrus reticulata
$G_3$	Rough lemon	Citrus jambhiri
$G_4$	Acid lime	Citrus aurantifolia
$G_5$	Nagpur Mandarin	Citrus reticulata
$G_6$	Sour orange	Citrus aurantium
$G_7$	Mosambi	Citrus sinensis
$G_8$	Cleopatra mandarin	Citrus reshni
$G_9$	Rangpur lime	Citrus limonia
$G_{10}$	Pani Jyamir	Citrus jambhiri
$G_{11}$	Grape fruit (Phokse)	Citrus paradisi
$G_{12}$	Pomelo	Citrus maxima
$G_{13}$	Kala Jyamir	Citrus jambhiri
$G_{14}$	Citron(Bimira)	Citrus medica

Table 2: Different concentration of aluminium

Sl. No	Treatments	Al concentration (µmol L <sup>-1</sup> )
1	$T_1$	0
2	$T_2$	50
3	$T_3$	100
4	$\mathrm{T}_4$	200
5	T <sub>5</sub>	400

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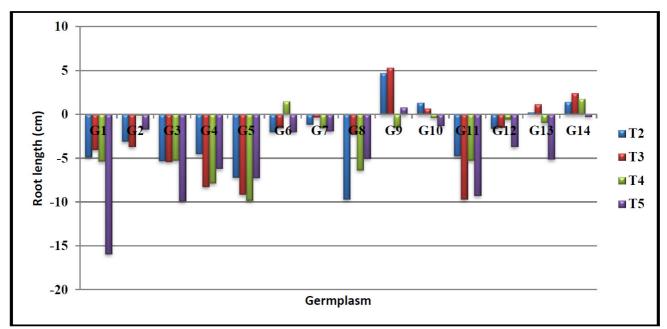


Fig 1: Reduction in root length of fourteen citrus germplasm under various aluminium concentration

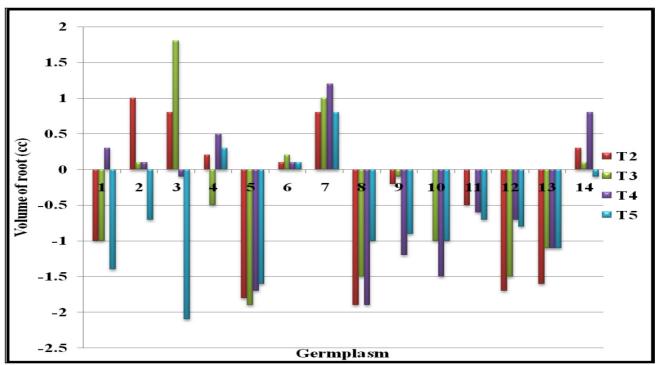


Fig 2: Reduction in root volume of fourteen citrus germplasm under various aluminium concentration

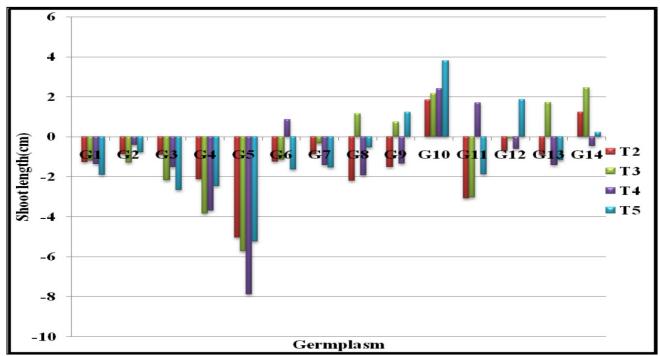
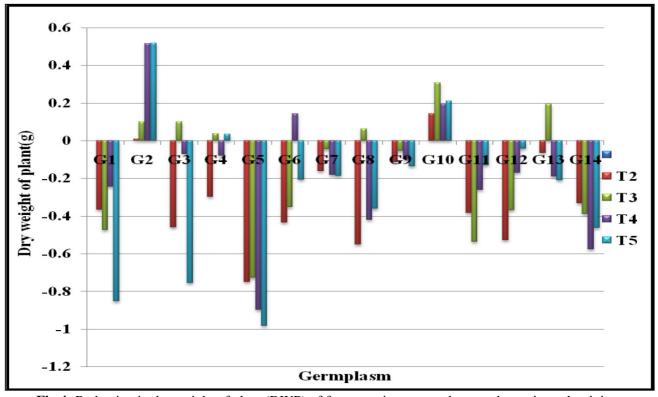


Fig 3: Reduction in shoot length of fourteen citrus germplasm under various aluminium concentration



**Fig 4:** Reduction in dry weight of plant (DWP) of fourteen citrus germplasm under various aluminium concentration

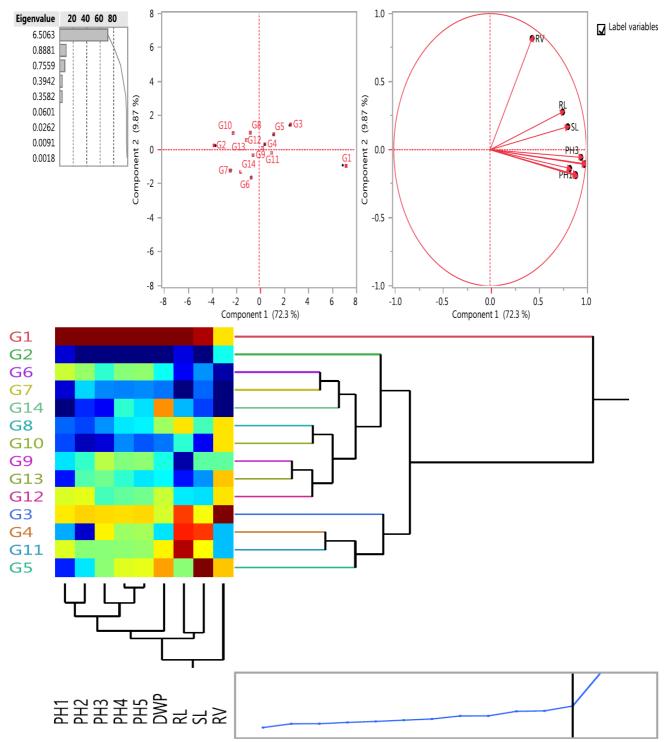


Fig 5: PCA and two way cluster analysis of citrus germplasm with plant parameter at A0

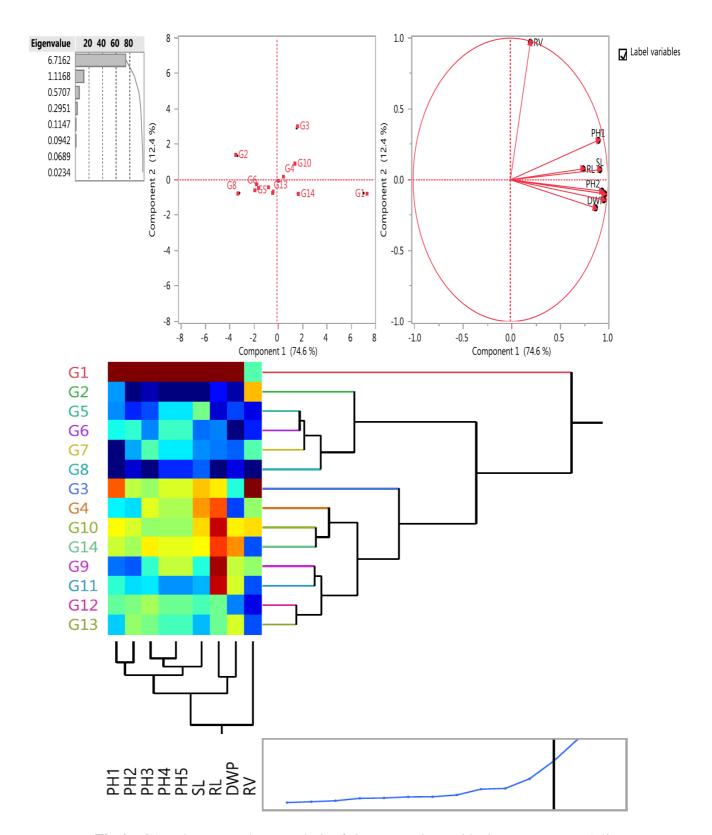


Fig 6: PCA and two way cluster analysis of citrus germplasm with plant parameter at A50

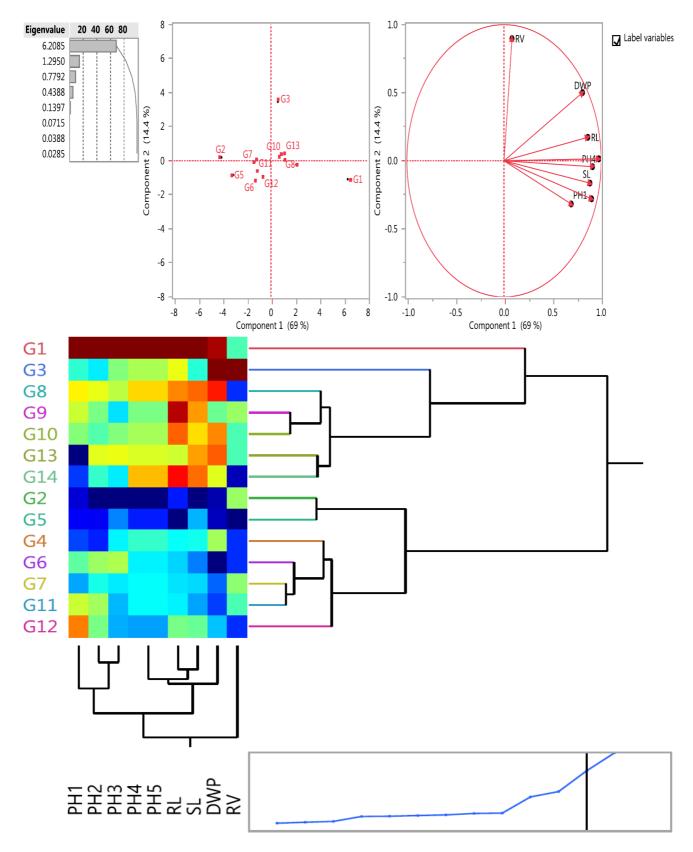


Fig 7: PCA and two way cluster analysis of citrus germplasm with plant parameter at A100

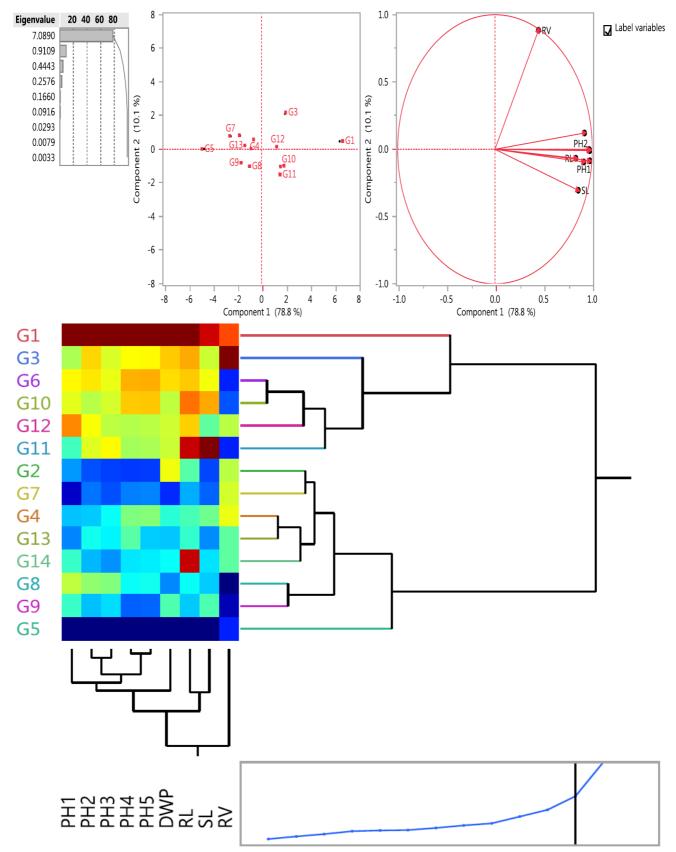


Fig 8: PCA and two way cluster analysis of citrus germplasm with plant parameter at A200

## Principal Components: on Correlations Summary Plots

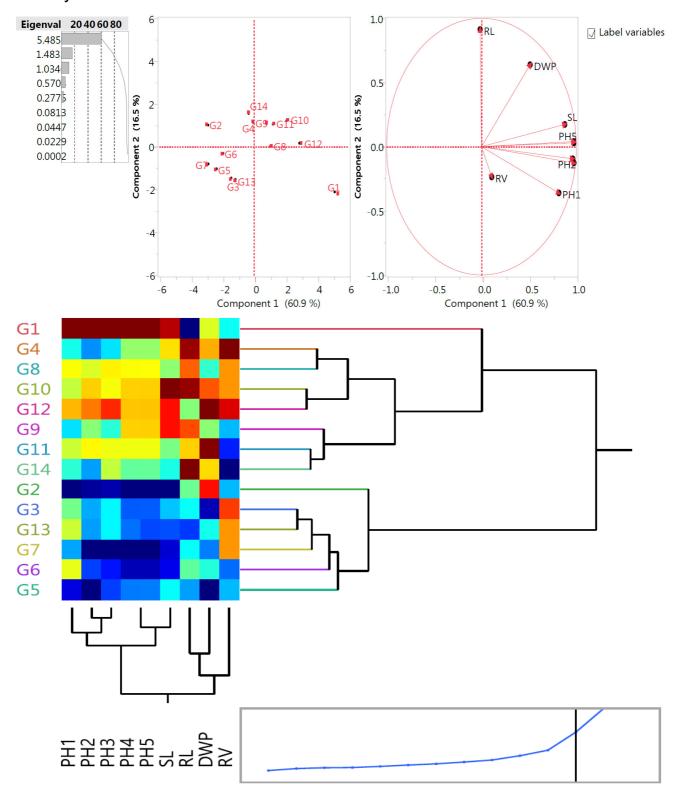


Fig 9: PCA and two way cluster analysis of citrus germplasm with plant parameter at A400

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#### References

- Batista, M.F., Moscheta, I.S., Bonato, C.M., Batista, A., Almeda, O.J.G. and Inoue, T.T. (2013). Aluminium in corn plants: Influence on growth and morpho-anatomy of root and leaf. *R. Bras. Ci. Solo.* 37 (1).
- Foy, C.D. (1984). Physiological effects of hydrogen, aluminium and manganese toxicities in acid soils. In: Adams, F. (Ed.). Soil Acidity and Limiting. Am. Soc. Agron. Madison. Wisconsin: 57–97.
- Guo, T.R., Zhang, G.P., Zhou, M.X., Lu, W.Y., Wu, H.P., Wu, H.B., Chen, J.X and Zhou, M.Z. (2003). Effect of aluminium on dry matter accumulation and aluminium and nutrients in barley differing in aluminium tolerance. *Plant Nutr. Fert. Sci.* **9**(3): 324-330.
- Haas, A.R.C. (1936). Phosphorus nutrition of citrus and beneficial effect of aluminium. Soil Sci. **42**:187-201.
- Lin, Z. and Myhre, D.L. (1991). Differential response of citrus rootstocks to aluminium levels in nutrient solutions: I. Plant growth. J. Plant Nutr. 14(11): 1223-1238.

- Magalhães, A.F.J. .(1987). Tolerance rootstock citrus aluminium. *J. Fr. cult.* **9** (3): 51-55.
- Nogueira, S.S., Nagai, V., Carelli, M.L.C. and Fahl, J.J. (1989). Comportamento de porta-enxertos de citros em presença de alumínio. *Pesq. Agropec. Bras.* **24**(6): 711-716.
- Pavan, M.A. and Bingham, F. (1982). Aluminum toxicity in coffee grown in nutrient solution. *Braz. Agri. Res.* 17(9): 1293-1302.
- Pereira, W. E., Siqueira de, D.L., Puiatti ,M., Martínez ,C, A., Salomão , L. C. C. and Cecon, P.R. (2003). Growth of citrus rootstocks under aluminium stress in hydroponics. *Sci. Agric.* **60**(1): 31-41.
- Santos, C.H., Son, H.G., Rodrigues, J. D. and Pine, S.Z. (1999a). Aluminium levels and the development of citrus rootstocks grown hydroponically: biometric parameters. *Sci. Agric.* **56** (4).
- Santos, C.H., Son, H.G., Rodrigues, J.D. and Pine, S.Z. (1999b). Aluminium levels and macro nutrients accumulation in rootstock citrus in growing hydroponic. *Sci.Agric.* **56** (4).
- Simon, L., Smalley, T. J., Jones, J. Benton. and Lasseigne, F.T. (1994). Aluminium toxicity in tomato. Part 1. Growth and mineral nutrition. J. Plant Nutr. 17(2-3): 293-306.
- Yokomizo, H. and Ishihara, M. (1973). Studies on the mineral nutrition of fruit trees by sand and water culture. I. In: Hokoku, K.S.A. (Ed.). Effects of composition of nutrient solution on the growth of Satsuma mandarin trees. 12: 29-77.